



Aerobic bacteria found in and around the carcasses of South American Sea Lions, *Otaria flavescens* (Carnivora, Otariidae), in Southern Brazil

ELIANE M. PEREIRA, DANIELA B. PEREIRA, JOANNA Z. ECHENIQUE; PATRICIA NASCENTE & ANA LUISA S. VALENTE*

Universidade Federal de Pelotas, Instituto de Biologia, Pelotas, RS, Brazil, 96160-000

*Corresponding author: schifinoval@hotmail.com

Abstract. Anaerobic bacteria were identified on the body surface and in the beach sand where carcasses of South American sea lion *Otaria flavescens* remain until decomposition. Swabs of the mouth, anus, back and skin of the fins and sand samples around the carcasses were collected from 10 adult specimens, in moderate decomposition state, along the coast of Southern Brazil. The swabs were seeded on blood agar, BHI, MacConkey, Rappaport and Salmonella-Shigella agar and incubated at 37°C for 24-48 hours. Aliquots of 25g of sand were diluted up to 10³ in 250 mL 0.1% peptone water. A 100 uL volume of each dilution was seeded on BHI agar and MacConkey agar and incubated. The colonies were submitted to Gram stain and identified by using the VITEK® automated system. Thirty-two species of bacteria were isolated and identified. *Proteus*, *Aeromonas*, *Kocuria* and *Sphingomonas* were found in 80% of the examined carcasses. *Proteus vulgaris* and *Sphingomonas paucimobilis* were the most common species in all samples (80%), followed by *Aeromonas salmonicida* (70%), *Kocuria rosea* (60%) and *Streptococcus thoralensis* (60%). Due to the presence of some bacteria with the potential to infect humans, both in the carcass and in sand nearby, the stranding site should be avoided by people in general.

Key words: pinniped, decomposition, microbial growth, zoonosis.

Resumo: Bactérias aeróbicas encontradas nas carcaças de leões-marinhos-sul-americanos, *Otaria flavescens* (Carnivora, Otariidae) e sedimento arenoso do entorno, no sul do Brasil.

Bactérias anaeróbicas foram identificadas na superfície do corpo e na areia da praia onde carcaças de leão-marinho sul-americano *Otaria flavescens* permanecem até sua decomposição. Amostras da boca, ânus, dorso e pele das nadadeiras e, amostras de areia ao redor das carcaças, foram coletadas em 10 espécimes adultos em estado de decomposição moderado, no litoral sul do Brasil. Os swabs foram semeados em ágar-sangue, BHI, MacConkey, Rappaport e ágar Salmonella-Shigella e incubados a 37°C por 24-48 horas. Alíquotas de 25g de areia foram diluídas até 10³ em 250 mL de água de peptona a 0,1%. Um volume de 100 uL de cada diluição foi semeado em ágar BHI e ágar MacConkey e incubado. As colônias foram submetidas à coloração de Gram e identificadas pelo sistema automatizado VITEK®. Trinta e duas espécies de bactérias foram isoladas e identificadas. *Proteus*, *Aeromonas*, *Kocuria* e *Sphingomonas* foram encontrados em 80% das carcaças. *Proteus vulgaris* e *Sphingomonas paucimobilis* foram as espécies mais comuns em todas as amostras (80%), seguidas por *Aeromonas salmonicida* (70%), *Kocuria rosea* (60%) e *Streptococcus thoralensis* (60%). Recomenda-se fortemente que, devido à presença de bactérias com algum potencial zoonótico, o acesso à praia próximo às carcaças de leões-marinhos-do-sul deva ser, de modo geral, evitado pelo público em geral.

Palavras-chave: Pinípede, decomposição, crescimento microbiano, zoonoses.

Introduction

Microorganisms are a significant component of the sand on beaches (Pinto & Oliveira 2011), which is an accumulating element of waste from different origins. This waste acts as a substrate for the presence of parasites and pathogens, which are dispersed in the environment (Lopes *et al.* 2008, Cicero *et al.* 2012). Microorganisms pathogenic to humans and fecal contamination indicators survive for long periods in beach sand due to the presence of organic matter (Vieira *et al.* 2007, Oliveira *et al.* 2015). As reported by the World Health Organization (WHO 1997), bacteria, fungi, parasites, and viruses have been observed in these conditions, and several genera and species are potentially pathogenic to humans through contact.

The proliferation of microorganisms is determined by the availability of organic matter in the sand, which can originate from plant or animal biomass (including decomposing bodies), animal feces and other waste of anthropogenic origin (Vieira *et al.* 2001, Kinzelman *et al.* 2003, WHO 2003). The vulnerability of beaches to pathogenic microorganism development is a public health hazard, because this environment is susceptible to contamination, particularly by helminths and protozoa present in animal feces, including *Ancylostoma* spp. and *Toxocara* spp. (Scaini 2003, Blazius *et al.* 2006, Cáceres *et al.* 2004). However, imbalances caused by this contamination, risks of transmission and even the identification of most pathogenic microorganisms are still unknown. The health and safety of recreational areas are primarily assessed by water quality, but studies have shown that the assessment of the microbiological quality of sand is also relevant (Alm *et al.* 2003, Maier *et al.* 2003, Lopes *et al.* 2008). In this context, sand can directly affect swimmers through causative agents of infectious diseases (Cetesb 2009, Passos *et al.* 2011).

Decaying animal carcasses that remain in coastal areas are an important source of organic matter and pose potential environmental and zoonotic risks. Given the topographical features and limited access to some areas, animal carcasses, such as those from seabirds, marine mammals, fish and even pets are often found along the coastline of Rio Grande do Sul, Brazil (ICMBio 2010). This probably happens because beaches are long and the stranding frequency of pinnipeds and cetacean carcasses is high, especially during the winter months (Pinedo *et al.* 1992). Marine mammals generally have a strong public appeal, as well as scientific and conservation interest. Close contact

with the body of these animals, whether by professionals (such as vets or biologists) or by ordinary people out of sheer curiosity, can pose risks to human health, including the transmission of zoonotic diseases and traumatic injuries (Waltzek *et al.* 2012).

Parasitic, fungal, and bacterial diseases have been reported in many marine animals (Dunn 1990, Dunn *et al.* 2001, Silva 1991; Cáceres *et al.* 2004; Pereira *et al.* 2020). In Brazil, the presence of *Vibrio* spp. has been identified in various species of marine mammals (Pereira *et al.* 2007), especially cetaceans. Furthermore, in this group of mammals, bacteria of the genera *Staphylococcus*, *Campylobacter*, *Aeromonas* and *Vibrio* have been isolated in the Franciscana dolphin *Pontoporia blainvillei* (Pereira *et al.* 2007, 2008, ICMBio 2010). In Sirenia, Delgado (2010) recorded the presence of *Leptospira* spp. in the Amazonian manatee (*Trichechus inunguis*).

In pinnipeds, a wide range of bacteria, including those of public health interest, have been isolated in various geographical areas (Gonzales *et al.* 2009, Carrasco *et al.* 2011). Among isolates is tuberculosis, which has been caused by *Mycobacterium bovis* or *Mycobacterium pinnipedii* in fur seals and sea lions in captivity (Forshaw & Phelps 1991); however, this disease has also been detected in *Arctocephalus australis* and *Otaria flavescens* in the coastal waters of Argentina (O'Reilly & Daborn 1995).

In southern Brazil, *O. flavescens* (Shaw, 1800), also known as the Patagonian sea lion, is endemic. This species is distributed along the rims of both coasts of South America, grouping together on rocky sites. In the Southwestern Atlantic Ocean, the northernmost grouping area of this species is southern Brazil, extending southwards as far as Tierra del Fuego, Argentina and continuing north along the Pacific as far as Peru (Vaz-Ferreira 1981, Rose *et al.* 1994). The species shows significant mortality rates on the southern coast of Rio Grande do Sul State, Brazil, where many animals arrive dead and others die locally, then undergoing decomposition processes (Pavanato *et al.* 2013). These carcasses are not collected and remain for several months under the action of tides, winds, and temperature variations until their total disappearance, by the consumption of scavengers or buried by sand. In the same geographical area, *O. flavescens* has been found dead on the beach and the specimens were highly parasitized by several species of helminths (Pereira *et al.* 2013).

Beach environmental quality has an increasing importance on environmental and public health grounds (Stewart *et al.* 2008) and beach sanitary quality should be monitored not only by the density of fecal coliforms present in waters. Thus, this study aimed to isolate and identify colonies of aerobic bacteria on the body surface of *O. flavescens* in moderate decomposition stage as well as in the soil surrounding the carcass deposition area to check the potential risks to public health.

Material and Methods

Carcass sampling: Swabs from the mouth, anus, back and skin of the fins of corpses of 10 adult specimens of *O. flavescens* found during systematic weekly field trips to the area between Torres (29 ° 20 'S) and Arroio do Chuí (33 ° 15'S), Rio Grande do Sul State, Brazil, were collected from October 2012 to December 2013. The distances between sampling points where the corpses were found did not follow a standard distribution, occurring as close as 300m and as far as 113km apart. Carcasses found in this period were at different stages of decomposition but only those in levels G2 or G3 (G2 – recently dead, fresh cadaver with complete integrity of eyes, and skin with complete fur coat, discrete fermentative process; G3 –relatively fresh cadaver, with complete fur coat, but with initial fermentative process causing abdominal distension, eyes dehydrated) were used, according to the protocol proposed by Geraci and Lounsbury (2005) adapted to pinnipeds. Therefore, the number of sampled sea lions does not represent an estimate of the mortality of this species in the studied area. The swabs were seeded on blood agar, BHI (Brain Heart Infusion), MacConkey, Rappaport and Salmonella-Shigella agar and incubated at 37° C for 24-48 hours.

Sand sampling: Four sand samples (125 cm³) taken from a depth of up to 5 cm and up to 10 cm from the mouth, anus and fins were collected for each carcass. Aliquots with 25g of each sand sample were diluted up to 10³ in 250 mL 0.1% peptone water. A 100 uL volume of each dilution was seeded on BHI agar and MacConkey agar and incubated at 37° C for 24-48 hours.

After growth, the respective bacterial colonies obtained were submitted to Gram stain and identified by using the VITEK® automated system; bacterial species were considered valid when identity was equal or up to 85%.

The categorization of each bacterial species' zoonotic potential was provided after extensive

bibliographical revision, considering as: NF (not found), when no record was found in human species; L (low), with just one record, in a sporadic clinical recording; E (Emerging) previous studies highlighting a new occurrence in more than one given case, considered as “emerging” in the article reviewed; M (medium), with relative occurrence in some geographical areas and in debilitated human patients; H (high), bacteria with recognized zoonotic potential and more than five records in human patients in different geographical areas.

Results

At least 32 species of bacteria from carcass and sand samples (n=80 samples) were isolated and identified (Table I). *Proteus vulgaris* and *Sphingomonas paucimobilis* were the most common species in all samples (80%), followed by *Aeromonas salmonicida* (70%), *Kocuria rosea* (60%) and *Streptococcus thoralensis* (60%). The percentage of identity of all isolated bacteria is presented in Table I.

Most of all identified bacteria were present in the carcasses, where bacteria of the genera *Aeromonas*, *Proteus*, *Kocuria* and *Sphingomonas* were found in 80% of the examined dead animals. Other genera including *Streptococcus* spp. (60%), *Staphylococcus* spp. (50%) and *Acinetobacter* (50%) were also found. *A. salmonicida* was identified in 80% of samples (Figure 1). The presence of *E. coli* was identified in 20% of carcasses and 10% of sand around the 10 carcasses of *O. flavescens*. The genera *Streptococcus* and *Staphylococcus* were isolated in 60% and 50% of the samples, respectively.

When bacterial growth in sand samples was contrasted to that found in carcasses, it was observed that only seven out of 32 identified species were present in both substrates: *A. salmonicida*, *A. haemolyticus*, *E. coli*, *P. vulgaris*, *S. warneri*, *S. intermedius* and *S. paucimobilis* (Table I, Figure 1).

Discussion

Several microbiological studies have pointed out long lists of species of aerobic bacteria that grow in cultures from samples of animal carcasses on beaches (Gomes 2014). However, the present study has extended the information on sand to monitor potential beach contamination and risks to public health. Even after finding a greater number of bacterial species in decomposing bodies than in the surrounding sand with less than a quarter sharing these resources, it is difficult to guarantee which is the primary source of contamination. These doubts

Table I. Aerobic bacteria isolated from *O. flavescens* carcasses and/or sand around carcasses in southern Brazil and their zoonotic potential. **NF (No record found as zoonosis), L (low), E (emerging), M (medium), H (high).** Bacteria present simultaneously in carcass and sand are typed in bold.

Bacterial Species	% Identities according to VITEK	Sand	Carcasses	Zoonotic potential	References
<i>Aeromonas salmonicida</i>	93	+	+	L,E	Altwegg & Geiss 1989
<i>Aeromonas hydrophila</i>	93	+	-	L,E	
<i>Acinetobacter haemolyticus</i>	90	+	+	L	Almasaudi 2016, Quinteira <i>et al.</i> 2007
<i>Acinetobacter ursingii</i>	93	+	-	L	Loubinoux <i>et al.</i> 2003
<i>Aloiococcus otitis</i>	99	-	+	L	Pereira <i>et al.</i> 2004
<i>Citrobacter freundii</i>	89	-	+	L	Badger <i>et al.</i> 1999
<i>Escherichia coli</i>	97	+	+	M	Kosilova <i>et al.</i> 2020
<i>Enterococcus faecium</i>	90	-	+	L	Steele <i>et al.</i> 2005
<i>Gemela sanguinis</i>	93	-	+	L, E	Purcell <i>et al.</i> 2001
<i>Kocuria rosea</i>	94	+	-	L, E	Leclercq <i>et al.</i> 1988
<i>Kocuria varians</i>	93	-	+	L, E	Kandi <i>et al.</i> 2016
<i>Kocuria kristinae</i>	89	+	-	L	Loong <i>et al.</i> 2017
<i>Leuconostoc mesenteroides</i> spp. <i>cremoris</i>	93	-	+	NF	Vagiakou-Voudris <i>et al.</i> 2002
<i>Pantoea</i> spp.	93	-	+	L	Panditrao & Panditrao 2017
<i>Pediococcus pentosaceus</i>	85	-	+	L	Ludlow <i>et al.</i> 2014
<i>Pasteurella pneumotropica</i>	89	-	+	L	Carriquiriborde <i>et al.</i> 2006
<i>Pseudomonas pseudoalcaligenes</i>	97	-	+	L	Hage <i>et al.</i> 2013
<i>Pseudomonas stutzeri</i>	98	+	-	L	Mattos <i>et al.</i> 2013
<i>Proteus vulgaris</i>	99	+	+	L	Burall <i>et al.</i> 2004
<i>Proteus mirabilis</i>	96	-	+	L	Burall <i>et al.</i> (2004
<i>Roseomonas gilardi</i>	99	-	+	L	Malini <i>et al.</i> 2017
<i>Rahnella aquatilis</i>	87	+	-	L	Martins <i>et al.</i> 2015
<i>Sphingomonas paucimobilis</i>	93	+	+	M	Baddour <i>et al.</i> 1985
<i>Staphylococcus saprophyticus</i>	86	-	+	L	Rodrigues <i>et al.</i> 1993
<i>Staphylococcus lentus</i>	99	+	-	L	Shaker <i>et al.</i> 2018
<i>Staphylococcus intermedius</i>	94	+	+	L	Llorca <i>et al.</i> 1992
<i>Serratia marcescens</i>	94	+	-	L	Del Peloso <i>et al.</i> 2010
<i>Streptococcus thoraltensis</i>	93	-	+	L	Dhotre <i>et al.</i> 2016
<i>Stenotrophomonas maltophilia</i>	97	-	+	L,E	Adsul <i>et al.</i> 2020
<i>Staphylococcus sciuri</i>	88	-	+	L	Hedin and Widerstrom 1998
<i>Staphylococcus warneri</i>	91	+	+	L	Incanni <i>et al.</i> 2010
<i>Shewanella algae</i>	98	+	-	L, E	Bertini <i>et al.</i> 2018

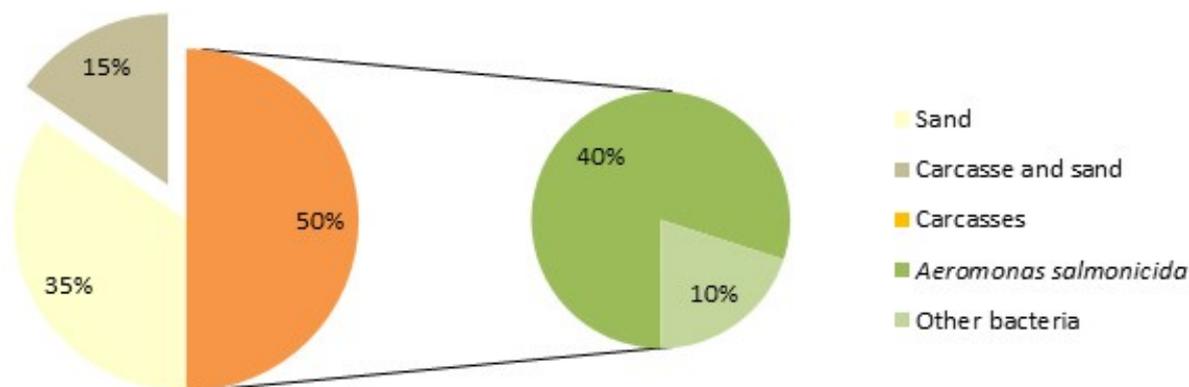


Figure 1. Proportional occurrence (%) of aerobic bacteria species (n=32) in *O. flavescens* carcasses or/and sand around carcasses in southern Brazil. Prevalence (%) is presented to *Aeromonas salmonicida*, considering the 40 samples from carcasses.

may be justified by the presence in the carcasses, even though less significant, of the 16 species which were found in the sand sediment. Considering the beach ecosystem where these carcasses were collected, high interactivity between abiotic and biotic factors in the grounding area was found. The time spent by the carcass on beaches attracts numerous scavengers, including seagulls, vultures, some Falconidae (caracaras), domestic and wild mammals, which may be responsible for the significant presence of organic matter and microorganisms on the site (Gomes 2014). On the other hand, bacteria associated with living sea lions and/or the decomposing tissues of their bodies may undergo washing by the constant action of tides, which, due to salinity, could limit sand colonization by some groups of microorganisms.

Contamination of sand on beaches is a risk to public health, and thus the recognition of each species or group of bacteria present and their potential for infection must always be taken into account. *E. coli* is one of the major constituents of intestinal microbiota in animals and, like thermotolerant coliforms, contaminates sand through feces (Murray 2006). Studies on the occurrence of *E. coli* associated with marine mammals are scarce (Salinas et al. 2010). Infection by *E. coli* is a disease

of increasing interest, both to human and animal health, as agents belonging to the family Enterobacteriaceae are responsible for significant incidences of pathological processes in humans and animals (Dho-Molin et al. 1999, Croxen et al. 2010).

Proteus vulgaris is naturally found in the digestive tract of humans and various other animal species (O'Hara et al. 2000). This bacterium is normally found in soil and water contaminated with fecal material and is an opportunistic pathogen known to cause infections in the urinary tract of human beings (Chow et al. 1979), as well as wound contamination (Pearson et al. 2008). Bacteria of the genus *Proteus* have been identified in the oral cavity of the South American fur seal *Arctocephalus australis* undergoing rehabilitation in southern Brazil (Vargas et al. 2008).

Sphingomonas paucimobilis is another microbial agent typically found in soil, which may occasionally cause septicemia, septic arthritis, and osteomyelitis in humans (Adley & Ryan 2010). Both bacteria were first associated with *O. flavescens* decomposition and may originate and be released during the fermentation process. Given their frequent occurrence, they stand out for their zoonotic potential through contact with mucous

membranes and damaged skin, which can represent a gateway for these opportunistic microorganisms.

Aeromonas salmonicida is a common pathogen that causes furunculosis and septicemia in fish (Kamble 2015). This organism usually infects homeothermic vertebrate animals that live in water at low temperatures. Until recently, it was thought that there was no bacterial growth at 37° C; however, a study on the microbiology of marine mammals has identified the presence of several species of *Aeromonas* in cetaceans on the Brazilian coast (Pereira *et al.* 2008). The same authors pointed out the presence of *A. caviae*, *A. hydrophila*, *A. veronii* biogroup *sobria*, *A. jandaei* and *A. media* in *O. flavescens*. The results of the present study agree with this previous record, because *A. salmonicida* was also the most representative bacterial genus, present in 80% of all samples from the carcasses of *O. flavescens*. The occurrence of *A. salmonicida* reported in this study is new for *O. flavescens* and, even if it cannot be guaranteed as associated with disease in this mammal, Tewari *et al.* (2014) reported the occurrence of this species associated with infection in humans. Species belonging to genus *Aeromonas* have been identified as causative agents of gastroenteritis and skin infections in humans; in addition, they are also considered important fish pathogens (KO *et al.* 2000, Scoglio *et al.* 2001). As *A. salmonicida* was identified both in carcasses and in sand around them, its occurrence seems to be causally linked to the presence of the corpse. The bacterium *A. hydrophila*, despite having been isolated from carcasses of species of cetaceans and even in live *O. flavescens* on the southern coast of Brazil (Pereira *et al.* 2008), was only identified in one sand sample of one animal (10%). This microbe is pathogenic and has been often isolated in samples of various origins, including water and animals, but has been mainly isolated from fish and mussels (Santos 1988, Pereira *et al.* 2004; Harnisz & Tucholski 2010).

Bacteria of the genus *Kocuria* are thought to cause emerging diseases in humans (Savini *et al.* 2010). Although the presence of species of this genus is known to be part of the skin microbiota of man, wild animals, and marine sediment (Kim *et al.* 2004), no records have been found on marine mammals. *Kocuria rosea* was found only in the sand around carcasses in 6 out of 10 animals examined, and its presence could not be associated with carcass decay. Regardless of its origin, this bacterium presents a contamination hazard to humans, since

there have been many records of infection in immuno-depressed individuals (Dunn *et al.* 2011).

According to Thornton *et al.* (1998), *Streptococcus* and *Staphylococcus* are commonly found on the skin of marine mammals. *Streptococcus* has subclinical signs, but may develop fever, depression, and various mastitis infections. In debilitated humans, it can cause respiratory symptoms, endocarditis, and skin lesions (Bisno *et al.* 1996). *Staphylococcus* infection in animals can cause skin lesions and pneumonia and eventually evolve to septicemia; in humans, it can cause abdominal pain, vomiting and diarrhea (Devriese *et al.* 2005). Among *Streptococcus*, *S. thoralensis* is a newly found species that was described by Devriese *et al.* (1997) in cultures originating from the intestine and genital tract of pigs. Although the genus *Streptococcus* now accommodates more than 60 species, relatively few of these streptococcal species have been recovered from marine animals. Indeed, only four species, *Streptococcus phocae* (Skaar *et al.* 1994) and *Streptococcus halichoeri* (Lawson *et al.* 2004) from seals, *Streptococcus iniae* recovered from freshwater dolphins and *Streptococcus marimammalium* from seals (Lawson *et al.* 2005) have been so far identified. In the present study, this species was mostly found in swabs collected directly from the rectum, which excludes the possibility of secondary contamination. This is therefore a new record of *S. thoralensis* for a marine mammal. Infections in humans are not known, but it is believed that the species usually colonizes the oral cavity (Dhotre *et al.* 2014).

Bacteria with high zoonotic potential, such as *Erysipelothrix insidiosa*, *Leptospira* spp., *Brucella* spp. and *Mycobacterium bovis*, which have already been isolated from pinnipeds (Blank *et al.* 2002), require specific methods for cultivation and identification which were not implemented in this study.

Given the findings of this study, including the presence of some microbial agents with the potential to infect humans, whether in carcasses or sand, access to the beach area next to the animal body may have negative implications for public health, and the use of protective equipment by professionals and researchers responsible for handling them is important.

Conclusions

A great diversity of aerobic bacteria colonizes the body surface of dead sea lions on temperate beaches in Southern Brazil. From the 32 bacterial

species identified, most from the sea lion bodies, only seven occurred simultaneously in the carcass and in the sand around it, suggesting low environmental contamination from carcasses. However, it is not yet possible to affirm which the primary source of colonization is, because biotic and abiotic factors interact in the stranding place, given the presence of other vertebrate and invertebrate animals, salinity variations, erosion and dehydration.,

From carcass surfaces, *Proteus*, *Aeromonas*, *Kocuria*, *Sphingomonas*, *Streptococcus*, *Staphylococcus* and *Acinetobacter* were the genera with greatest prevalence. Essentially, considering all samples (body plus sand samples = 80 samples) the most frequent bacteria were *A. salmonicida*, *Kocuria rosea* and *Streptococcus thoralensis*, which accounted for between 60-70% of bacteria found. They do not have a high zoonotic potential except in immunosuppressed individuals, for which sporadic records were found in the literature. As an important recommendation, due to the presence of some bacteria with the potential to infect humans, both in the carcasses of South American sea lions and in sand nearby, the stranding site should be avoided by people in general.

Acknowledgments

We are grateful to E. Secchi, responsible for the Laboratório de Megafauna at the Instituto de Oceanografia da Universidade Federal do Rio Grande, for allowing us to participate in routine beach trips to collect samples. We thank J. Mendes for help during the microbiology lab procedures. Thanks are also due to the two anonymous reviewers that contributed to making this manuscript clearer.

Conflict of Interest: The authors declare that they have no conflict of interest.

Statement on the welfare of animals: The present study used only animals that had died of natural causes. The samplings were performed in cooperation with the Laboratório de Megafauna at the Instituto de Oceanografia da Universidade do Rio Grande. Permission was provided by the **Brazilian Institute of Environment and Renewable Natural Resources – SISBIO** - under license number 16586- 2 and registration number 2031900.

References

- Adsul, N. M., Panigrahi, V., Acharya, S. Kalra, K. L. & R. S. Chahal. 2020. *Stenotrophomonas maltophilia* spondylodiscitis following lumbar microdiscectomy mimicking a cotton granuloma: A case report and literature review. *Surgical Neurology International*, 25: 11- 28.
- Alm, E. W., Burke, J., & Spain, A. 2003. Fecal indicator bacteria are abundant in wet sand at freshwater beaches. *Water Research Philadelphia*, 37: 3978 – 3982.
- Almasaudi, S. 2016. *Acinetobacter* spp. as nosocomial pathogens: Epidemiology and resistance features. *Saudi Journal of Biological Sciences*. 25. 10.1016/j.sjbs.2016.02.009.
- Altwegg, M. & Geiss, H. K. 1989. *Aeromonas* as a human pathogen. *Critical Reviews in Microbiology*, 16:253-86
- Baddour, L. M, Kraus, A. P., Jr., & Smalley, D. L. 1985. Peritonitis due to *Pseudomonas paucimobilis* during ambulatory peritoneal dialysis. *Southern Medical Journal*, 78: 366.
- Badger, J. L., Stins, M. F. & Kim, K. S. 1999. *Citrobacter freundii* invades and replicates in human brain microvascular endothelial cells. *Infection and immunity*, 67: 4208–4215.
- Blank, O., Retamal, P., Abalos, P. & Torres, D. 2002. Detección de anticuerpos Anti-*Brucella* en focas de Weddell (*Leptonychotes weddellii*) de Cabo Shirref Antártica. *Archivos de Medicina Veterinaria*, 34: 117-122.
- Blazius, R. D., Silva, O. S., Kauling, A. L. Rodrigues, D. F. P., & Lima, M. C. 2006. Contaminação da areia da praia do Balneário de Laguna, SC, por *Ancylostoma* spp., e *Toxocara* spp. em amostras fecais de cães e gatos. *Arquivos Catarinense de Medicina*, 35:3.
- Bisno, A. L., & Stevens, D. L. 1996. Streptococcal infections of skin and soft tissues. *New England Journal of Medicine*, 25: 240-244.
- Burall, L. S., Harro, J. M., Li, X., Lockett, C. V., Himpl, S. D., Hebel, J. R., Johnson, D. E., & Mobley, H. L. 2004. *Proteus mirabilis* genes that contribute to pathogenesis of urinary tract infection: identification of 25 signature-tagged mutants attenuated at least 100-fold. *Infection and immunity*, 72: 2922–2938.
- Carrasco, S. E., Burek, K. A., & Beckmen, K. B. 2011. Aerobic oral and rectal bacteria of free-ranging steller sea lion pups and juveniles (*Eumetopias jubatus*) in Alaska. *Journal of Wildlife Diseases*, 47: 807–820.
- Carriquiriborde, M ; Milocco, S ; Príncipi, G & Cagliada, P & Carbone, C. 2006. *Pasteurella pneumotropica* causa la regresión de tumores humanos trasplantados en ratones

- imunodeficientes. *Medicina* (Buenos Aires), 66: 242-244
- CETESB. 2009. Relatório de Qualidade das Praias Litorâneas: no Estado de São Paulo. São Paulo: Cetesb,.
- Croxen, M. A., & Finlay, B. B. 2010. Molecular mechanisms of *Escherichia coli* pathogenicity. *Nature*, 8: 20-38.
- Cicero, L. H., Quiñones, E. M., Cunico, P., & Santos, C. L. 2012. Contaminação das areias de praias do Brasil por agentes patológicos. *Revista Ceciliansa*, 4: 44-49.
- Chow, A. W., Taylor, P. R., Yoshikawa, T., & Guze, L. B. 1979. A nosocomiae outbreak of infection due to multiply resistant *Proteus mirabilis*: role of intestinal colonization as a major reservoir. *Journal of Infectious Diseases*, 130: 621-627.
- Delgado, P. M., Gennari, S. M., Madeira, A. M. B. N., & Soares, I. S. 2011. *Detecção sorológica de infecção por Toxoplasma gondii e Leptospira spp. em peixes-bois (Trichechus inunguis) de dois centros de preservação da Amazônia brasileira*. Universidade de São Paulo.
- Del Peloso, P. F, Barros, M. F. L., & Santos, F. A. 2010. Sepsis por *Serratia marcescens* KPC. *Jornal Brasileiro de Patologia e Medicina Laboratorial*, 46: 365-367. <https://dx.doi.org/10.1590/S1676-24442010000500004>
- Devriese, L. A., Vancanneyt, M., Baele, M., Vanechoutte, M., De Graef, E., Snauwaert, C., Cleenwerck, I., Dawyndt, P., Swings, J., Decostere, A., & Haesebrouck, F. 2005. *Staphylococcus pseudintermedius* sp. a coagulase-positive species from animals. *International Journal of Systematic and Evolutionary Microbiology*, 55: 1569–1573.
- Devriese, L. A., Pot, B., Vandamme, P., Kersters, K., Collins, M. D., Alvarez, N., Haesebrouck, F., & Hommez, J. 1997. *Streptococcus hyovaginalis* sp. nov. and *Streptococcus thoralensis* sp. nov., from the genital tract of sows. *International Journal of Systematic Bacteriology*, 47(4):1073 e 1077.
- Dho-Moulin, M., & Fairbrother, J. M. 1999. Avian pathogenic *Escherichia coli* (APEC). *Veterinary Research* 30:299-316.
- Dhotre, S. V., Mehetre, G. T., Dharme, M. S., Suryawanshi, N. M & Nagoba, B. S. 2014. Isolation of *Streptococcus tigurinus* novel member of *Streptococcus mitis* group from a case of periodontitis. *FEMS Microbiology Letters*, 357: 131-135.
- Dhotre, S., Suryawanshi, N., Nagoba, B. & Selkar, S. 2016. Rare and unusual isolates of viridans streptococci from the human oral cavity. *Indian Journal Pathological Microbiology*, 59: 47-9.
- Dunn, J. 1990. Bacterial and mycotic diseases of cetaceans and pinnipeds. In: Dierauf, L.A. (Ed.), *Handbook of Marine Mammal Medicine Health, Disease and Rehabilitation*. CRC Press, USA, pp.73-87.
- Dunn, J. L., Buck, J. D., & Robeck, T. R. 2001. Bacterial diseases of cetaceans and pinnipeds. In: Dierauf L.A., Gulland F.M.D. (eds.) *CRC handbook of marine mammal medicine*, 2nd ed. CRC Press, Boca Raton, FL. pp. 312–314.
- Dunn, R., Bares, S., & David, M. Z. 2011. Central venous catheter-related bacteremia caused by *Kocuria kristinae*: Case report and review of the literature. *Annals of Clinical Microbiology and Antimicrobials*, 10:31
- Forshaw, D., & Phelps, G. R. 1991. Tuberculosis in a captive colony of pinnipeds. *Journal of Wildlife Diseases* 27: 288-295.
- Geraci, J. R. & Lounsbury, V. J. 2005. *Marine Mammals Ashore: A Field Guide for Strandings*. National Aquarium in Baltimore, Baltimore. 2ª Ed. pp.177-188.
- González, M. J., Villanueva, M. P., Latif, F., Fernández, F. & Fernández, H. 2009. Aislamiento de *Plesiomonas shigelloides* y *Aeromonas veronii* biotipo *sobria* en heces de lobo marino común sudamericano, *Otaria flavescens* (Shaw, 1800). *Revista de Biología Marina y Oceanografía*, 44:763-765.
- Gomes, S., Macedo, M. R. P., Pesenti, T., Pereira, D. I. B., Cirne, M. P. & Müller, G. 2014. Isolamento de *Salmonella enterica* subsp. *diarizonae* em *Calidris fuscicollis* (Aves: Scolopacidae) no Rio Grande do Sul, Brasil. *Ornithologia*, 8:43-45.
- Hage, J. E., Schoch, P. E. & Cunha, B. A. 2013. *Pseudomonas pseudoalcaligenes* peritoneal dialysis-associated peritonitis. *Perit Dial Int.*;33(2):223-224. doi:10.3747/pdi.2012.00112
- Harnisz, M., & Tucholski, S. 2010. Microbial quality of common carp and pikeperch fingerlings cultured in a pond fed treated wastewater. *Ecological Engineering*, 36:466-470.
- Hedin, G. & Widerstrom, M. 1998. Endocarditis due to *Staphylococcus sciuri*. *European Journal of Clinical and Microbiology and Infectious Diseases*,17:673-675

- Incáni, R. N., Hernández, M., Cortez, J., González, M. E., & Dorel, S. Y. 2010. *Staphylococcus warneri* meningitis in a patient with *Strongyloides stercoralis* hyperinfection and lymphoma: first report of a case. *Revista do Instituto de Medicina Tropical de São Paulo*, 52: 169-170.
- Kamble, R. 2015. *Aeromonas salmonicida* furunculosis in an adult male. *International journal of Current Microbiology Applied Science*, 4:59-63.
- Kandi, V., Palange, P., Vaish, R., Bhatti, A. B., Kale, V., Kandi, M. R., & Bhoomagiri, M. R. 2016. Emerging Bacterial Infection: Identification and Clinical Significance of *Kocuria* Species. *Cureus*, 8(8). doi:10.7759/cureus.731
- Kinzelman, J., Clement, N. G., Jackson, E., Gradus, S., & Bagley, R. 2003. Enterococci as indicators of Lake Michigan recreational water quality: comparison of two methodologies and their impacts on public health regulatory events. *Applied Environmental Microbiology*, 69:92-96.
- Kim, S. B., Nedashkovskaya, O. I., Mikhailov, V. V., Han, S. K., Kim, K. O., Rhee, M. S. & Bae, K. S. 2004. *Kocuria marina* sp. nov., a novel actinobacterium isolated from marine sediment. *International Journal of Systematic Evolutionary Microbiology*, 54: 1617–1620.
- Ko, W. C., Lee, H. C., Chuang, Y. C., Liu, C. C., & Wu, J. J. 2000. Clinical features and therapeutic implications of 104 episodes of monomicrobial *Aeromonas bacteremia*. *Journal of Infectology*, 40: 267-273.
- Kosilova, O. Y., Vovk, O. O., Katelevska, N. M., Osolodchenko, T. P., Ponomarenko, S. V. & Vdovichenko, V. Y. 2020. Study of pathogenic factors of *E.coli* isolated from patients with peritonitis. *Wiad Lek.* 73:78-82.
- Lawson, P. A., Foster, G., Falsen, E., Davison, N. & Collins, M. D. 2004. *Streptococcus halichoeri* sp. Nov., isolated from grey seals (*Halichoerus grypus*). *International Journal of Systematic Evolutionary Microbiology*, 54:1753-1756.
- Lawson, P.A., Foster, G., Falsen, E., & Collins, M. D. 2005. *Streptococcus marimammalium* sp. nov., isolated from seals. *International Journal of Systematic and Evolutionary Microbiology*, 55: 271–274
- Leclercq, R., Derlot, E., Duval, J. & Courvalin, P. 1988. Plasmid-mediated resistance to vancomycin and teicoplanin in *Enterococcus faecium*. *New England Journal of Medicine*, 319: 157-161
- Llorca, I., S. Gago, J. Sanmartin, & Sanchez, R. 1992. Endocarditis infecciosa por *Staphylococcus intermedius* en paciente infectado por VIH. *Enf. Infect. Microbiol. Clin.* 10:317–318.
- Lopes, F. W. A., Magalhães Júnior, A. P. & Pereira, J. A. A. 2008. Avaliação da qualidade das águas e condições de balneabilidade na bacia do Ribeirão de Carrancas-MG. *Revista Brasileira de Recursos Hídricos* 13:111-112.
- Loubinoux, J., Mihaila-Amrouche, L., LeFleche, A., Pigne, E., Huchon, G., Grimont, P. A. D. & Bouvet, A. 2003. Bacteremia Caused by *Acinetobacter ursingii*. *Journal of Clinical Microbiology*, 41: 1337-1338
- Ludlow, S. & Pasikhova, Y. 2014. A Case Report of *Pediococcus pentosaceus* Bacteremia Successfully Treated with Daptomycin. *Infectious Disease in Clinical Practice*, 75. 45. 10.1097/IPC.0b013e318281d905.
- Maier, L. M., Rezende, K. C. R. Vieira, V. D. R., Carvalho, C. R., & Seixas, J. T. F. 2003. Avaliação dos fungos e bactérias patogênicas nas areias de duas praias de baixo hidrodinamismo e alta ocupação humana no município do Rio de Janeiro. *Augustus* 8: 24.
- Malini, S., Goh, B. L. & Lim, T. S. 2016. A Rare Case of *Roseomonas gilardii* Peritonitis in a Patient on Continuous Ambulatory Peritoneal Dialysis. *Perit Dial Int.*, 36:578.
- Martins, W., Carvalhaes, C. G., Cayô, R., Gales, A. C. & Pignatari, A. C. 2015. Co-transmission of *Rahnella aquatilis* between hospitalized patients The Brazilian Journal of Infectious Diseases 19:648-650
- Mattos, F. B., Agostini, F. S., Mattos, M. B. & Batista, D. M. P. 2012. Úlcera de córnea por *Pseudomonas stutzeri*. *Revista Brasileira de Oftalmologia*, 71:111-114.
- Murray, P. R., Rosenthal, K. S. & Pfaller, M. A. 2006. *Microbiologia Médica*. 5 a Edição Rio de Janeiro: Elsevier.
- Oliveira, B. S., Rédua, C. R. O., Souza, Y. S. & Fernandes, E. S. 2015. Ocorrência de helmintos eggs and larvae ground of squares, day care and public schools in the city of Luziânia-Goiás, Brazil. *Scientific Electronic Archives* 8:2.
- O'hara, C. M., Brenner, F. W. & Miller, J. M. 2000. Classification, identification, a clinical significance of *Proteus*, Providencia, and

- Morganella*. *Clinical Microbiology Review*, 13:534-546.
- O'Reilly, L. M. & Daborn, C. J. 1995. The epidemiology of *Mycobacterium bovis* infections in animals and man: a review. *Tuber Lung Dis. Aug* 1:1-46.
- Panditrao, M. & Panditrao M. 2018. *Pantoea dispersa*: Is it the Next Emerging "Monster" in our Intensive Care Units? A Case Report and Review of Literature. *Anesth Essays Res* 12: 963-966. doi: 10.4103/aer.AER_147_18
- Passos, C. T., Silva, A. P. R., Braga, A. R. C., Sanzo, A.V.L., & Kalil, S.J. 2011 Variação Sazonal da contaminação por coliformes na areia e água da praia do Cassino, Rio Grande, RS. *Arquivos de Ciências do Mar*, 44:21 – 26.
- Pavanato, H., Silva, K. G., Estima, S. C., Monteiro, D. S., & Kinas, P. G. 2013. Occupancy dynamics of South American Sea-Lions in Brazilian Haul-outs. *Brazilian Journal of Biology*, 73:855-862.
- Pearson, M., Sebahia, M., Churcher, C., Quail, A. A., Seshasayee, A. S., Luscombe, N. M., Abdellah, Z., Arrosmith, C., Atkin, B., Chillingworth, T., Hauser, H., Jagels, K., Moule, S., Mungai, S., Norbertszak, H., Rabinowitsch, E., Walker, D., Whithead, S., Tomson, N. R., Rather, P. N., Parkhill, J., & Mobley, H. L. T. 2008 Complete genome sequence of uropathogenic *Proteus mirabilis*, a master of both adherence and motility. *Journal of Bacteriology*, 190: 4027-4037.
- Pereira, C. S., Possas, C. A., Viana, C. M. & Rodrigues, D. P. 2004a. *Aeromonas* spp. e *Plesiomonas shigelloides* isoladas a partir de mexilhões (*Perna perna*) in natura e pré-cozidos no Rio de Janeiro, RJ. *Ciênc. Tecnol. Aliment* 24: 562-566.
- Pereira, M., Rotta, B., Cantarelli, V., Pereira, D. & Costa, S. S. 2004b. Prevalência elevada do *Alloiooccus otitidis* na otite média com efusão através da PCR simultânea. *Revista Brasileira de Otorrinolaringologia*, 70(2), 217-224. <https://doi.org/10.1590/S0034-72992004000200012>
- Pereira, C. T., Amorim, S. D., Santos, A. F. M., Siciliano, S., Moreno, I. M. B., Ott, P. H. & Rodrigues, D. P. 2007. *Vibrio* spp. isolados de mamíferos marinhos capturados na região litorânea do sudeste ao sul do Brasil. *Pesquisa Veterinária Brasileira*, 27:81-83.
- Pereira, C. S., Amorim, S. D., Santos, A. F. M., Siciliano, S., Moreno, I. M. B., Ott, P. H. & Rodrigues, D. P. 2008. *Plesiomonas shigelloides* and *Aeromonadaceae* family pathogens isolated from marine mammals of southern and southeastern Brazilian coast. *Brazilian Journal of Microbiology*, 39: 749-755
- Pereira, E. M., Müller, G., Secchi, E., Pereira Jr, J. & Valente, A. L. S. 2013. Digenetic Trematodes in South American Sea Lions from Southern Brazilian Waters. *Journal of Parasitology*, 99:910-913.
- Pereira, E. M., Mendes, J. E., Echenique, J. V. Z., Brayer, D. I., Valente, A. L. S. 2020. Fungal colonization on body surfaces of dead South American sea lions on sandy beaches, and an evaluation of risk of contamination to humans at Southern Brazil. *Brazilian Journal of Development*, 6: 53799-53812.
- Pinedo, M. C., Rosas, F. C. & Marmontel, M. 1992. Cetáceos e Pinípedes do Brasil. Uma revisão dos registros e guia para identificação das espécies. UNEP/FUA, Manaus, Brasil. p.213.
- Pinto, A. B. & Oliveira, A. J. F. C. 2011. Diversity of indicator microorganisms in the evaluation of sea recreational beach sand fecal contamination: current knowledge and perspectives. *O mundo da saúde* 35:105-114
- Purcell, L. K., Finley, J. P., Chen, R., Lovgren, M. & Halperin, S.A. 2001. *Gemella* species endocarditis in a child. *Canadian Journal Infectious Diseases*, 12:317-320. doi:10.1155/2001/960734
- Quinteira, S., Grosso, F., Ramos, H. & Peixe, L. 2007. "Molecular Epidemiology of Imipenem-Resistant *Acinetobacter haemolyticus* and *Acinetobacter baumannii* Isolates Carrying Plasmid-Mediated OXA-40 from a Portuguese Hospital" (PDF). *Antimicrobial Agents and Chemotherapy*. 51:3465–3466.
- Ryan, M. P. & Adley, C. C. 2010. *Sphingomonas paucimobilis*: a persistent Gram-negative nosocomial infectious organism. *Journal of Hospital Infection*, 75:153-7.
- Rodrigues, M. A. V., Reis, C., & dos Santos, R. A. 1993. *Staphylococcus saprophyticus* em infecções do trato urinário masculino e feminino. *Journal of Tropical Pathology*, 22(2). <https://doi.org/10.5216/rpt.v22i2.20058>.
- Santos, Y., Toranzo, A. E., Barja, J.L., Nieto, T. P., & Villa, T. G. 1988. Virulence properties and enterotoxin production of *Aeromonas* strains isolated from fish. *Infectology and Immunology* 56:3285-3293.

- Salinas, P., Moraga, R., Santander E. & Sielfeld, W. 2010. Presencia de cepas diarreogénicas de *Escherichia coli* y estudio de genes de virulencia en aislados desde fecas de dos poblaciones de lobo marino común, *Otaria flavescens* en el norte de Chile. *Revista de Biología Marina y Oceanografía* 45: 153-158.
- Savini, V., Catavittello, C., Masciarelli, G., Astolfi, D., Balbinot, A., Bianco, A., Febbo, F., D'Amario, C. & D'Antonio, D. 2010. Drug sensitivity and clinical impact of members of the genus *Kocuria*. *Journal of Medical Microbiology* 59:1395–1402.
- Scaini, C. J. 2003. Environmental contamination by helminth eggs and larvae in dog feces from central area of Cassino beach, Rio Grande do Sul. *Revista da Sociedade Brasileira de Medicina Tropical*, 36: 617-619.
- Scoglio, M. E., Di Pietro, A., Picerno, I., Delia, S., Mauro, A. & Lagana P. 2001. Virulence factors in *Vibrios* and *Aeromonads* isolated from seafood. *New Microbiology*, 24: 273-80.
- Skaar, L., Gaustad, P., Tonjum, T., Holm, B. & Stenwig, H. 1994. *Streptococcus phocae* sp. Nov., a new species isolated from clinical specimens os seals. *Internal Journal of Bacteriology*, 44: 646-650.
- Shaker M. N., Hmdan T. A. & Issa A. H. 2018. Isolation and Diagnosis of *Staphylococcus lentus* from different operation theater hospitals. *The Journal of Science Medical Research*, 2 : 177-181
- Steele, C. M., Brown, R. N & Botzler, R. G. 2005. Prevalences of zoonotic bacteria among seabirds in rehabilitation centers along the pacific coast of California and Washington, USA *Journal of Wildlife Diseases*, 41: 735–744
- Stewart, J. R., Gast, R. J., Fujioka, R. S., Solo-Gabriele, H. M., Meschke, J. S., & Zettler, L. A. 2008. The coastal environment and human health: microbial indicators, pathogens, sentinels, and reservoirs. *Environmental Health*, 7 (Suppl 2): S3.
- Thornton, S. M., Nolan, S. & Gulland, F. M. D. 1998. Bacterial Isolates from California Sea Lions (*Zalophus californianus*), Harbor Seals (*Phoca vitulina*), and Northern Elephant Seals (*Mirounga angustirostris*) Admitted to a Rehabilitation Center along the Central California Coast, 1994-1995. *Journal of Zoo and Wildlife Medicine* 29: 171-176.
- Vaz-Ferreira, R. 1981. South American sea lion *Otaria flavescens* (Shaw) In: Ridgway S. & Harrison R. Handbook of Marine Mammals. New York: Academic Press (1): 39-66.
- Vieira, R. H. S. F., Rodrigues, D. P., Manezes, E. A., Evangelista, N. S. S., Reis, E. M. F., Barreto, L. M. & Gonçalves, F. A. 2001. Microbial contamination of sand from major beaches in Fortaleza, Ceará, Brazil. *Brazilian Journal of Microbiology* 32: 77-80.
- Vieira, R. H. S. F., Oliveira, A. C. N. & Sousa, O.V. 2007. Monitoramento microbiológico das águas e areias das praias do Meireles e do Futuro, Fortaleza /Ceará. *Boletim Técnico Científico da Cepnor, Belém* 7(1): 17 – 26.
- WHO, 2015 (World Health Organization) Regional Office for Europe. Development of WHO Guidelines for Safe Recreational Water Environments. Report on a WHO Expert Consultation. St Helier, Jersey, United Kingdom, 23 -30 May 1997. Disponível em <<http://www.euro.who.int/document/e58484.pdf>> Acesso em:12 fev. 2015.
- Vagiakou-Voudris E., Mylona-Petropoulou, D., Kalogeropoulou, E., Chantzis, A., Chini, S., Tsiodra, P., & Malamou-Lada, E. 2002. "Multiple Liver Abscesses Associated with Bacteremia due to *Leuconostoc lactis*". *Scandinavian Journal of Infectious Diseases*, 34: 766–7.
- Vargas, R. R., Albano, A. P. N. & Silva Filho, R.P. 2008. Flora bacteriana da cavidade oral e nasal de lobo-marinho-do-sul (*Arctocephalus australis*) em centro de reabilitação. *Anais do 35° CONBRAVET*, 19 a 22 de outubro 2008. Gramado, RS, Brasil.
- Waltzek, T. B., Cortés Hinjosa, G., Wellehan Jr., J. F. X. & Gray, G. C. 2012. and Public Health. Marine Mammal Zoonoses: A Review of Disease Manifestations. *Zoonoses and Public Health* 59(8): 521–535.

Received: September 2020

Accepted: January 2021

Published: March 2021